

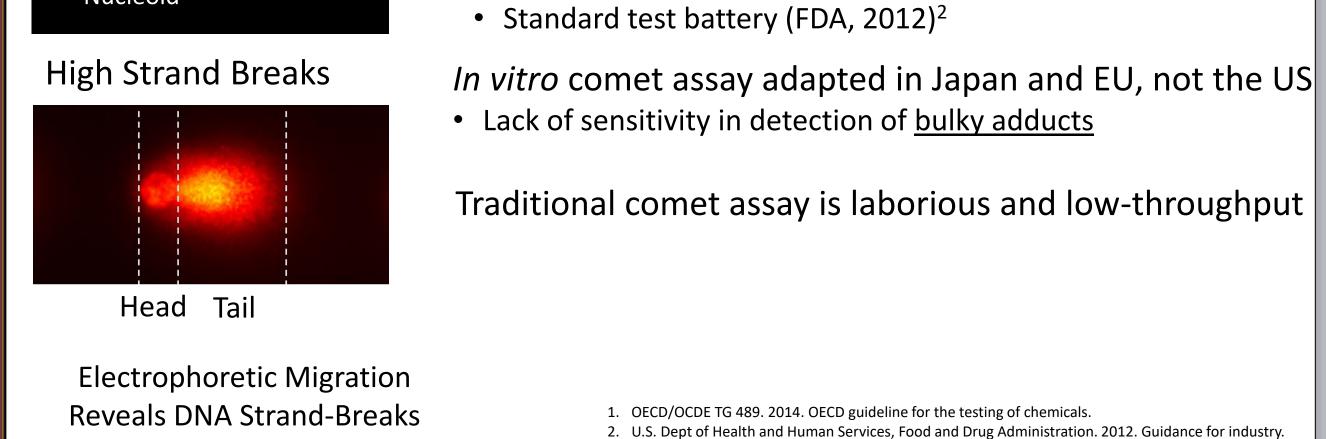


#### ABSTRACT

Rational: The CometChip assay is a well established, high throughput, in-vitro method for the detection

# No Damage Nucleoid

Modified from Wood D et al. 2010 PNAS **107**-10008-10013



guidelines

## Exploiting AI To Improve DNA Damage Detection and Quantification.

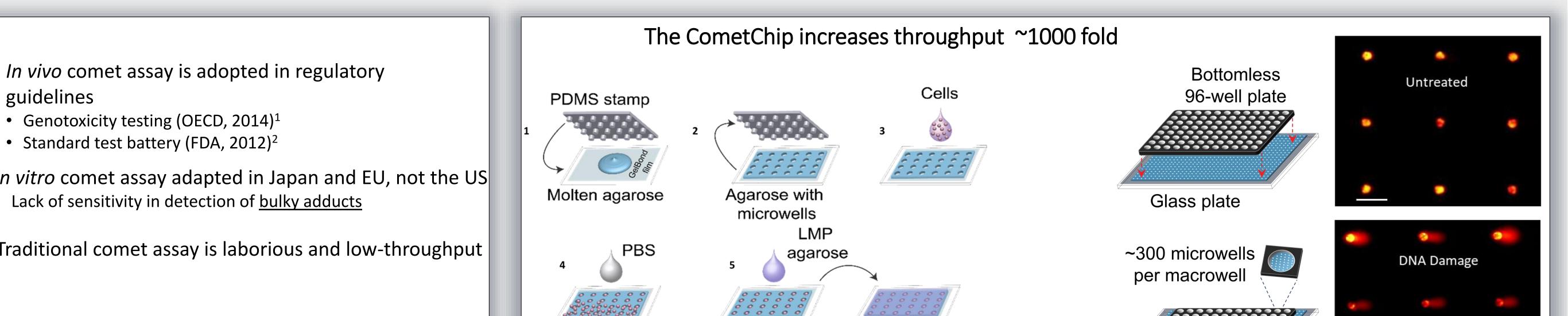
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Micropatterned

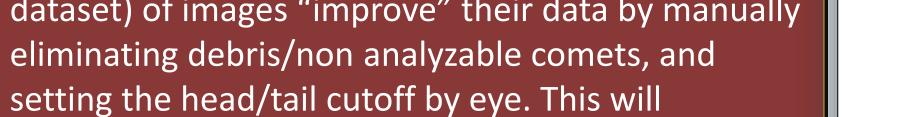
### The Comet Assay

• Genotoxicity testing (OECD, 2014)<sup>1</sup>

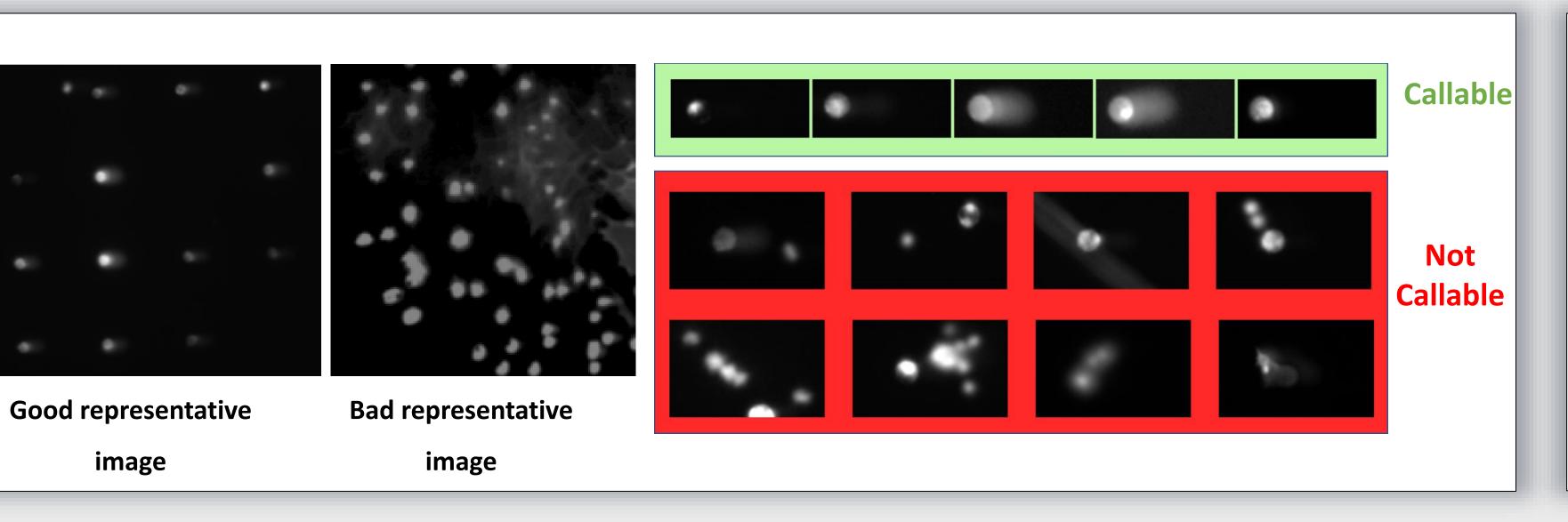
### The CometChip Platform



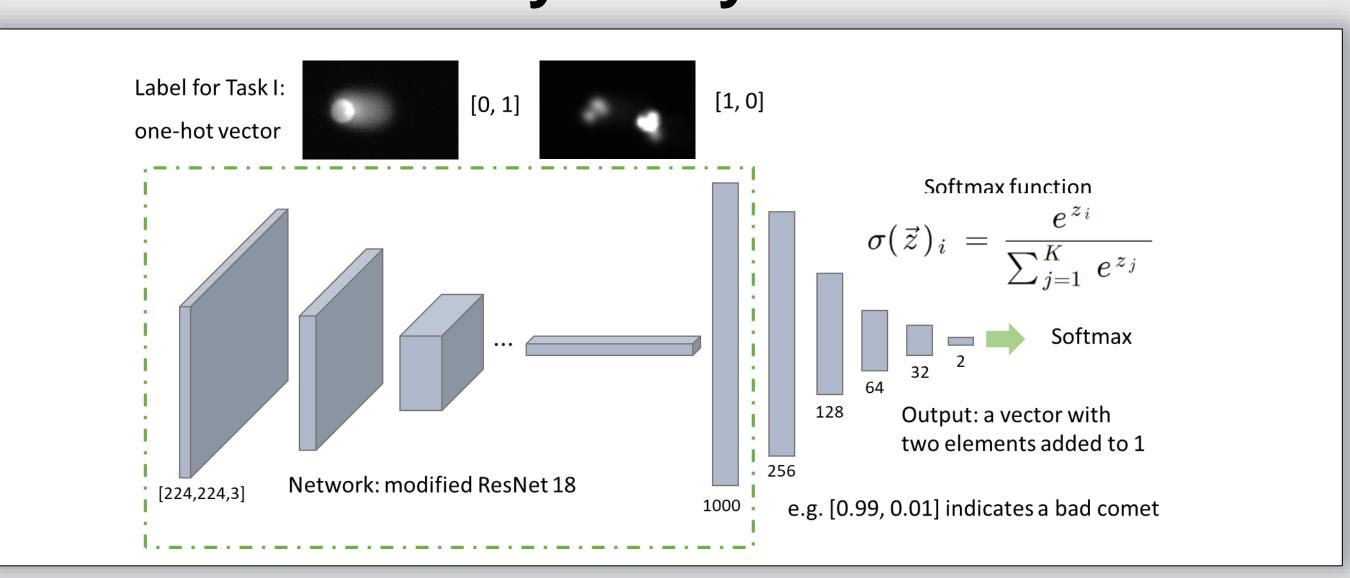
and quantification of DNA damage in cells based on the Comet assay, developed in 1984. Current commercially available analysis software is expensive, proprietary, and requires manual input from the user. Manipulation of data analysis by eye increases the amount of time necessary to analyze each image dataset and introduces bias that can affect DNA damage quantification. This projects aims to use machine learning to train an image analysis algorithm that can ultimately be used to fully automate image analysis. Fluorescent images of electrophoresed comets must be analyzed to calculate the extent of DNA damage. Expensive commercially available analysis software is used to determine the main parameter which is the cutoff between the "head" (intact DNA with low mobility) and the "tail" (damaged DNA with higher mobility) of the comet. The percentage of total DNA in the tail represents the amount of damage. Current software cannot determine the quality of individual comets, and often the software places the cutoff line far from the correct location forcing users to painstakingly manually sift through hundreds (a very small dataset) to hundreds of thousands (a large dataset) of images "improve" their data by manually eliminating debris/non analyzable comets, and

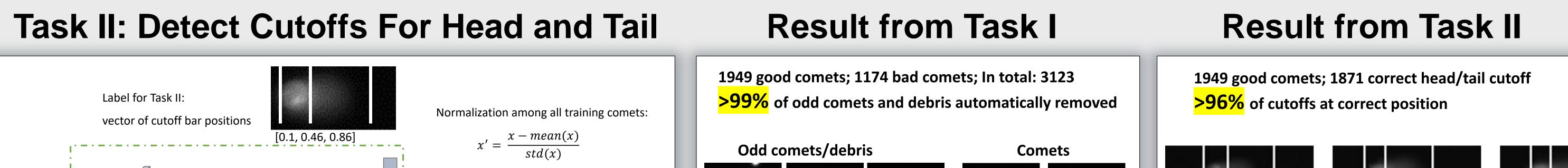






## **Task I: Identify Analyzable Comets**





eliminate the possibility of manual input from users, greatly increasing throughput (by orders of magnitude, in some cases) and providing objective and consistent analysis parameters.

**Comet vs CometChip Assay: the** Comet assay relied on randomly patterned cells on an agarose coated glass slide, this led to significant overlapping when overloaded, and few cells on the plate when underloaded, creating high degrees of variability in data quality between slides, as well as a low number of images per slide even under ideal loading conditions. The CometChip assay exploits micropatterned micro-wells in agarose that can be separated into the "macro" wells of a typical 96 well plate.

#### **Current progress**

machine learning has been applied to both automation steps, leading to a 99.7% (9997/1000) screening of debris or ununiform comets in the first step, and an over 95% correct cutoff determination in the second step. We will soon be incorporating these into a free to use program that will be compared with existing



#### **Saturation Issues and Comet Tail Over-Counting Good comets** ••• S10 ·••· S20 ·••· S50 ← S20@1.6 Barely Visible -- S20@0.8 0.25253686 ••• S100 -- S20@0.1 --- S20@0.025 0.088202566 uper Saturated 0.00020573652 1,000 1e+04 0.1100 1,000 100 1e+04 Exposure time (ms) Exposure (ms) **Determining guidelines for Pixel Saturation Conclusions and Significance**

Saturated pixels in raw image vs DNA damage quantification - S10 sat pixels

• The CometChip platform is robust and high-throughput, will soon have a free and equally high throughput analysis software • Machine learning is capable of producing very high accuracy in results without input from user. • Ideal exposure times have been empirically determined and guideline set for the effects of image pixel saturation which will be accounted for in finished product. • Once completed, this open-source, free software will remove any fiscal hurdles for CometChip image analysis, and drastically increase throughput and fidelity of DNA damage quantification using CometChip.



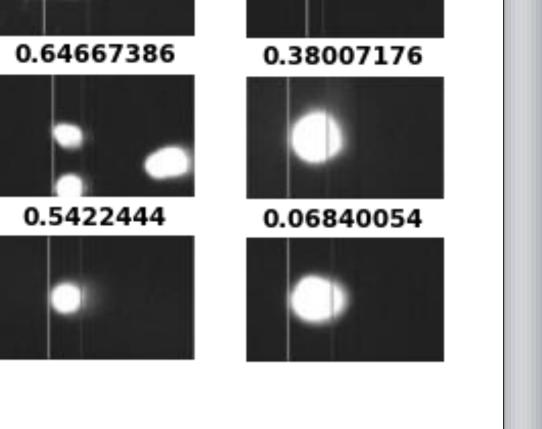


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0.38254032

**Bad comets** 

0.5580731

0.76701766

0.6555367

0.6015375

